



## Concise review

# Impact of apolipoprotein E genetic polymorphisms on liver disease: An essential review

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## ABSTRACT

Cirrhosis is an advanced stage of liver disease, compromising liver function with systemic health implications and poor quality of life. Hepatitis C virus (HCV) infection and alcoholic liver disease are the main causes of this pathology. However, since genetic factors may play a large role in the progression and severity of liver disease, and as apolipoprotein E (apoE) has been recognised to be mainly synthesised in the liver, apoE polymorphism studies are important to better understand the causal mechanisms in liver diseases. In this review, we summarise up-to-date studies addressing how apoE polymorphisms influence liver cirrhosis and liver transplantation outcomes and potential protective mechanisms. Although more clinical studies are needed to support these findings, the apoE ε4 allele seems to be protective against the progression of liver cirrhosis in the majority of aetiologies and the postoperative serum apoE phenotype of the transplanted subject receptors was converted to that of the donor, indicating that >90% of apoE in plasma is synthesised in the hepatic system.

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## 1. Introduction

Cirrhosis is the result of the advanced stage of liver disease, resulting from the replacement of the functional liver architecture with non-functional fibrotic tissue, being responsible for important health problems worldwide [1,2]. Infections by hepatitis C virus (HCV) and alcoholic liver disease are the main causes of liver cirrhosis [3,4]. However, this liver pathology can also be caused by other aetiologies such as metabolic diseases, hereditary (hemochromatosis, Wilson's disease, Alpha-1-antitrypsin deficiency), non-alcoholic steatohepatitis (NASH), autoimmune hepatitis, Budd-Chiari syndrome, primary biliary cirrhosis (PBC), and primary sclerosing cholangitis [1,2]. Therapeutic options include liver transplantation, which is the definitive treatment in patients with end-stage liver cirrhosis, improving the quality of life as well as the longevity of this population [1].

Human apolipoprotein E (apoE) consists of a 34 kDa glycoprotein, containing 299 amino acid residues, which is an important protein component of very low density lipoproteins (VLDL) and a ligand for the low density lipoprotein receptor (LDL-R) [5–7].

ApoE is synthesised mainly in the liver, but also in the spleen, brain, kidney, lungs, adrenal gland, monocyte-macrophage, muscle tissues, central and peripheral nervous system. It has important actions in neuronal repair, in the regulation of lipid homeostasis, and in the transport and metabolism of triglycerides and cholesterol. It also has anti-inflammatory functions, skewing the pro-inflammatory macrophagic phenotype M1 to the anti-inflammatory M2 and decreasing the synthesis of interleukin-2 (IL-2), as well as roles in immunomodulatory activities such as the activation and proliferation of T lymphocytes [7–10].

Some studies have documented the association between apoE isoforms and diseases such as Alzheimer's disease, atherosclerosis, liver disease caused by HCV, human immunodeficiency virus (HIV) infection, HIV-associated dementia, pulmonary tuberculosis, childhood diarrhoea and herpes simplex virus infection [5,7,11,12].

ApoE was also evidenced in an experimental model as a sensitive marker in the graft function of transplanted hepatocytes, which is important since hepatocyte transplantation has been

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emphasised as a promising treatment for patients with liver or metabolic diseases and acute liver failure [13].

It is well documented that genetic factors play a key role in the severity and progression of liver diseases. Therefore, we set out to review the role of apoE polymorphisms in conditioning the natural history of pre- and post-transplant liver disease, as well as its association with the development of liver fibrosis and response to therapies.

## 2. Polymorphism and different apolipoprotein e isoforms

The human apoE gene is polymorphic. In humans, this polymorphism is responsible for different apoE isoforms, due to amino acid substitutions at position 112 and 158 [5,14,15]. There are three common alleles of the apoE gene, which is located on chromosome 19 (19q13); these are called  $\epsilon 2$ ,  $\epsilon 3$  and  $\epsilon 4$ , with six possible genotypes:  $\epsilon 2/\epsilon 2$ ,  $\epsilon 2/\epsilon 3$ ,  $\epsilon 2/\epsilon 4$ ,  $\epsilon 3/\epsilon 3$ ,  $\epsilon 3/\epsilon 4$ , and  $\epsilon 4/\epsilon 4$ . The  $\epsilon 3$  allele is the most frequent isoform, accounting for 70–80% of alleles worldwide, encoding the apoE3 isoform with a cysteine residue at position 112 and an arginine residue at position 158. The  $\epsilon 4$  allele encodes the apoE4 isoform, with an arginine residue at both positions 112 and 158, whereas the  $\epsilon 2$  allele has a point mutation causing the replacement of arginine for cysteine at position 158. These account for 10–15% (for the  $\epsilon 4$  allele) and 5–10% (for the  $\epsilon 2$  allele) of the alleles worldwide [5,7,9].

Previous studies have shown that the apoE  $\epsilon 4$  allele may increase plasma triglyceride levels and decrease the levels of high density lipoprotein (HDL) cholesterol. In addition, it is associated with higher low density lipoprotein (LDL) cholesterol levels compared to the  $\epsilon 3$  isoform. However, the  $\epsilon 2$  isoform has a close association with hypertriglyceridaemia and hypocholesterolaemia [16]. Noteworthy, ~15% of  $\epsilon 2/\epsilon 2$  homozygous patients may develop familial dysbetalipoproteinaemia (also known as type III hyperlipoproteinaemia). This leads to hypercholesterolaemia and hypertriglyceridaemia and may be associated with obesity and insulin resistance [17,18].

## 3. Apolipoprotein E and orthotopic liver transplantation

Orthotopic liver transplantation is currently the most appropriate treatment for end stage liver disease, and may also influence the synthesis and degradation of genetically polymorphic-encoded plasma proteins. Thus, as apoE is a polymorphic protein in humans, some studies have already reported that it is possible to detect and quantify changes in this protein in patients who have undergone liver transplantation [9,15].

Linton et al. [15] performed a 29-patient sample study and reported that the postoperative serum apoE phenotype of the receptor was converted to that of the donor. This indicates that >90% of apoE in the plasma is synthesised in the hepatic system. On the other hand, there was no change in the apoE phenotype of cerebrospinal fluid (CSF) from the donor to the receptor phenotype after hepatic transplantation, indicating that most of the apoE in CSF is synthesised locally and not derived from plasma. Kraft et al. [9] also emphasised that more than 90% of apoE in humans is of hepatic origin, since the new apoE phenotype following liver transplantation corresponded to that of the donor organ.

In experimental models, serum apoE is a sensitive marker with which to monitor the functioning and survival of grafts of transplanted hepatocytes. These results are of significant importance, since hepatocyte transplantation may be a promising technique for patients with metabolic liver disease or acute liver failure [13]. In addition, by affecting lipoprotein metabolism, apoE polymorphisms may modulate the recurrence of HCV in individuals undergoing liver transplantation [19].

## 4. Apolipoprotein E, hepatitis B and C viruses and hepatocellular carcinoma

### 4.1. Hepatitis B virus

Hepatitis B virus (HBV) infection is still very common (and worrying) in developing countries. Its aetiological agent is a DNA virus, hepatovirus of the family *Hepadnaviridae*; in particular, the long-standing infection can lead to liver failure, cirrhosis and hepatocellular carcinoma (HCC) [20–22].

Previous studies have documented the association between apoE genotypes and viral diseases, including herpes simplex virus (HSV), HIV, HCV, and HBV [21,23,24]. It has been documented that the  $\epsilon 3$  allele of the apoE was frequent in patients with progressive HBV-related liver cirrhosis [25].

In addition, Shen et al. [21] enrolled 40 healthy volunteers and 199 patients with HBV, active hepatitis, severe hepatitis, cirrhosis and HCC, and demonstrated that serum levels of apoE  $\epsilon 3$  progressively increased with disease severity (HBV carriers developed hepatitis, followed by cirrhosis and then ultimately HCC). The  $\epsilon 3$  allele and  $\epsilon 3/\epsilon 3$  genotype were the most prevalent in all subgroups. Furthermore, progressive serum elevation of interleukin-6 (IL-6) and a gradual decrease of IL-2 were associated with disease progression and severity, while apoE serum levels were positively correlated with serum IL-6 (but not with IL-2).

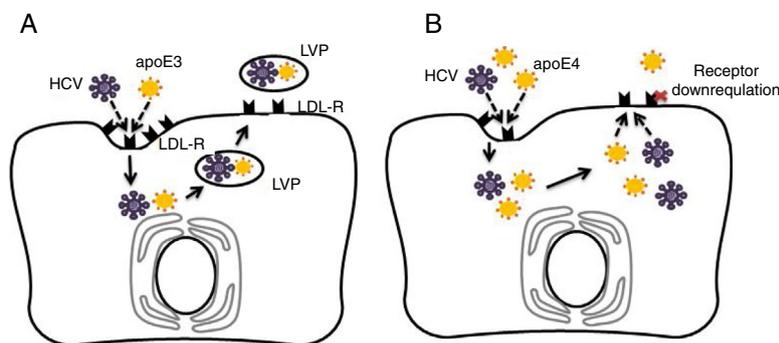
### 4.2. Hepatitis C virus

The hepatitis C virus is a flavivirus and hepatotropic RNA virus, which can cause acute and chronic hepatitis C, as well as liver cirrhosis and hepatocellular carcinoma in humans [26]. However, there is a recent suggestion of a correlation between low cholesterol levels and infection by viruses C. Authors have reported that the mechanisms of HCV, HSV and HIV infection resemble each other, as all of these viruses compete with apoE to bind to cell receptors [23,27].

ApoE polymorphisms may be an important tool for monitoring the progression of fibrosis in patients with hepatitis C and normal alanine aminotransferase levels, as there may be competition mechanisms for viral entry and replication [27]. In other reports, HCV synthesised in individuals carriers of the apoE  $\epsilon 2$  allele is associated with a low risk of infection and rapid elimination [28].

Pioneering studies by Wozniak et al. [24] have shown that the apoE  $\epsilon 4$  allele was significantly more frequent in patients with chronic hepatitis C and mild liver disease compared to those with severe disease, indicating that the  $\epsilon 4$  allele may be protective against liver injury due to HCV. Mueller et al. [29] also showed a significant low frequency of the apoE  $\epsilon 4$  allele in patients diagnosed with chronic HCV infection, suggesting a protective action of this allele.

In the study by Price et al. [30] a significant lower frequency of both the  $\epsilon 2$  and  $\epsilon 4$  alleles was associated with a reduced infection risk in patients with HCV. Mueller et al. [31] also suggested a protective role of the  $\epsilon 4$  allele and a higher risk of persistent HCV infection in  $\epsilon 3$  allele carriers in chronically HCV-infected patients. Furthermore, a higher frequency of the  $\epsilon 4$  allele was found among the non-cirrhotic chronic hepatitis C patients, supporting that the  $\epsilon 4$  allele is protective against HCV infection [32]. In another study with 996 chronically HCV-infected patients, the apoE  $\epsilon 4$  allele was poorly represented [31]. In addition, the same authors documented reduced viral loads in patients with the apoE  $\epsilon 4$  allele who were chronically infected with HCV genotype type 1 [29], suggesting that this allele has a protective effect against HCV infection.



**Fig. 1.** Comparison of apolipoprotein E3 (apoE3) and apolipoprotein E4 (apoE4) effects on hepatitis C virus (HCV) infection in hepatocytes. (A) Normal cycle of HCV entry in hepatocytes via low density lipoprotein receptor (LDL-R). HCV competes with apoE particles (apoE3 in this case) for binding of LDL-R, ending in the formation of lipo-viral-particle (LVP), which is important for virus infectivity. (B) ApoE4 proposed protection against HCV entry. In addition of apoE and virus competition for cell entry via LDL-R, the apoE4 carriers would show down-regulated LDL-receptors in hepatocytes and less viral capsulating.

#### 4.3. Potential mechanisms for apoE4 protection against hepatitis C virus infection

As discussed previously in our review, apoE4 has been suggested as a protective factor against HCV infection [33,34]. The entry of HCV into human hepatocytes is a multistep mechanism in which various host factors are involved, including LDL-R and heparan sulphate proteoglycans (HSPGs). The lipoviral particle (LVP), which is important for viral infectivity, initially binds LDL-R and HSPGs through apoE [35–37]. It has been recognised that the LDL-R is down-regulated in apoE4 carriers [23]. Thus, in apoE  $\epsilon$ 4 patients, the virus entry in hepatocytes may be reduced [10]. Fig. 1 depicts the potential protective mechanism of apoE  $\epsilon$ 4 against HCV in hepatocytes.

ApoE would be the only specific factor for the production of infectious HCV particles; therefore, apoE could influence a late stage of virus infectivity after viral capsid envelopment, being essential for viral cell-to-cell transmission [38]. One hypothesis is that apoE  $\epsilon$ 4 indirectly affects LDL receptor expression by increasing the binding and internalisation of lipoproteins by apoE  $\epsilon$ 4 [31,39]. On the other hand, apoE  $\epsilon$ 4-induced hyperbetalipoproteinemia could directly interfere with uptake of the LDL-R mediated virus due to forced competition between free betalipoproteins and virus-lipoprotein particles for local LDL receptors [40]. Thus, the apoE4-related functional properties in lipid metabolism (and negative regulation of liver LDL receptors) may provide an explanation (corroborated with epidemiological evidence) of how apoE4 carriers are protected against HCV infection.

#### 4.4. Hepatocellular carcinoma

Hepatocellular carcinoma (HCC) is the most frequent primary liver cancer [41], and is mostly found in patients with liver cirrhosis, mainly due to infection by the hepatitis B and C viruses. Thus, as cirrhosis is the main risk factor associated with HCC, liver transplantation became the main treatment of this tumour [41–44].

Some studies have shown that apoE has antioxidant actions and is increased in malignant tumours such as gastric, prostate, ovarian and HCC, mainly due to the oxidative stress generated by the tumour cells [45–47]. Yokoyama et al. [48] found that apoE levels in tumour tissues were significantly higher than in normal non-tumour tissues in 88% of patients, without any increase in the plasma, and that apoE may be a histological marker for HCC. They also showed that 72.7% of patients with HCC were  $\epsilon$ 3/ $\epsilon$ 3.

Ahn et al. [25] found that the apoE plasma level was significantly higher in the group of patients with hepatic cirrhosis and the HCC group. They also observed that the  $\epsilon$ 3 allele and the  $\epsilon$ 3/ $\epsilon$ 3 genotype were the most frequent in both groups and that  $\epsilon$ 4 allele was

the one that presented the lower probability of developing liver cirrhosis.

#### 5. Apolipoprotein E and primary biliary cirrhosis

Primary biliary cirrhosis (PBC) is an autoimmune cholestatic liver disease due to inflammation of small intrahepatic biliary ducts, which can result in fibrosis and cirrhosis of the liver in some patients; liver transplantation is the last therapeutic approach [49–51]. ApoE polymorphisms can modify the severity of PBC, acting on intestinal absorption and the excretion of bile salts. The phenotype may have an influence on the pathogenesis of PBC and also act on the disease response during treatment [50,51].

Corpechot et al. [50] reported that the  $\epsilon$ 3 allele and the  $\epsilon$ 3/ $\epsilon$ 3 genotype were the most frequent in PBC patients; however, there was no difference in both the disease and French control population, suggesting that  $\epsilon$ 3 is not a risk factor for the onset of PBC in the French population. Furthermore, after PBC treatment with ursodeoxycholic acid, among  $\epsilon$ 4 carriers, the liver enzymes did not return to baseline, suggesting a poor response to therapy in these patients. Conversely, in the study by Vuoristo et al. [51], the  $\epsilon$ 2 allele was seen significantly more often in patients with PBC, while the  $\epsilon$ 4 allele carriers showed better liver enzyme tests when treated with ursodeoxycholic acid. More studies with larger numbers are needed to better dissect these contradictory results.

#### 6. Apolipoprotein E and alcoholic liver cirrhosis

Alcoholic liver cirrhosis due to chronic alcohol abuse has been a longstanding worldwide health problem, resulting in worrisome increasing in morbidity and mortality [52]. The incidence of the risk of death is higher in cirrhotic patients than the risk of developing liver cirrhosis, which indicates that low or moderate alcohol consumption is not related to significant increases in the risk of developing cirrhosis; however, this risk tends to increase exponentially from excessive alcohol intake [53]. In addition, not every individual who consumes alcohol regularly will develop alcoholic cirrhosis, as genetic and environmental factors also contribute to the development of this condition. However, it is important to emphasise that alcohol has an important effect on lipid metabolism, leading to hypertriglyceridaemia, hypercholesterolaemia and changes in lipoproteins [54,55].

However, previous studies have documented the influence of apoE polymorphisms in patients who developed hepatic cirrhosis due to alcohol [23,55]. In an experimental model, the authors observed that chronic long-term ingestion of ethanol in hepatic alcohol dehydrogenase-deficient deer mice led to the infiltration of T lymphocytes into the liver and hepatic steatosis. There was also

**Table 1**  
ApoE polymorphism studies on liver cirrhosis in humans.

References	Aetiology	Studies characteristics	Outcomes	Allele frequencies
Mueller et al. 2009 [4]	HCV	701 patients with chronic HCV infection, 523 healthy controls and 283 non-HCV patients.	Risk of chronic HCV infection.	↓ $\epsilon 4$
Wozniak et al. 2002 [24]	HCV	156 anti-HCV positive patients and 104 non-HCV infected.	Risk of severe disease.	↓ $\epsilon 4$
Fabris et al. 2011 [27]	HCV	128 Caucasians, PNALT and HCV carriers. A subgroup of 116 patients with absent or minimal fibrosis was follow up for 10 years.	Rapid progression of fibrosis.	↑ $\epsilon 3/\epsilon 3$ ↓ $\epsilon 2/\epsilon 2 = \epsilon 4/\epsilon 4$
Mueller et al. 2003 [29]	HCV	Retrospective analysis, treatment response of 506 chronically HCV-infected patients.	Reduced HCV virus genotype 1.	↑ $\epsilon 4$
Price et al. 2006 [30]	HCV	420 Northern European patients with evidence of exposure to HCV were compared with 288 healthy controls.	Risk of HCV infection.	↓ $\epsilon 2 = \epsilon 4$ ↓ $\epsilon 2/\epsilon 2$
Mueller et al. 2016 [31]	HCV	701 patients with chronic HCV infection and 295 additional patients with chronic HCV infection, 179 patients with a history of spontaneous HCV clearance; 283 patients with non-HCV-associated chronic liver disease and 2234 healthy controls.	HCV infection: Protective risk	↑ $\epsilon 4$ ↑ $\epsilon 3$
Teama et al. 2016 [32]	HCV	Case-control study: 80 chronic hepatitis C patients (40 cirrhotic and 40 non cirrhotic), and 40 healthy controls.	Risk of severe disease.	↓ $\epsilon 4$
Shen et al. 2015 [21]	HCC and HBV	40 healthy volunteers and 199 patients (30 HBV carriers, 60 with active hepatitis, 12 with severe hepatitis, 58 with HBV and liver cirrhosis, and 39 with HCC).	Progressive with disease severity.	↑ $\epsilon 3$ , ↑ $\epsilon 3/\epsilon 3$ ↓ $\epsilon 2$ , ↓ $\epsilon 2/\epsilon 4$
Ahn et al. 2012 [25]	HCC and HBV	Case-control study: 47 healthy controls, 156 patients (50 with HBV and liver cirrhosis, and 59 with HCC and liver cirrhosis).	Risk of developing liver cirrhosis.	↑ $\epsilon 3$ , ↑ $\epsilon 3/\epsilon 3$ ↓ $\epsilon 4$ , ↓ $\epsilon 4/\epsilon 4$ ↑ $\epsilon 3$
Yokoyama et al. 2006 [48]	HCC	17 patients (14 HCV carriers and 3 HBV carriers).	Production in HCC tissues.	↑ $\epsilon 3$
Corpechot et al. 2001 [50]	PBC	Retrospective cohort: 72 PBC patients were treatment for 4 years with ursodeoxycholic acid (12–15 mg/kg/day) and thereafter compared with 1808 healthy French controls.	No susceptibility to PBC. Risk of severe disease and poor response to treatment.	$\epsilon 2 = \epsilon 3 = \epsilon 4$ ↑ $\epsilon 4$
Vuoristo et al. 1997 [51]	PBC	88 patients with PBC were randomised to ursodeoxycholic acid, placebo treatment or colchicine for 2 years.	PBC severity. Expression of PBC and response to treatment.	$\epsilon 2 = \epsilon 3 = \epsilon 4$ ↑ $\epsilon 2$ , ↓ $\epsilon 4$
Iron et al. 1994 [54]	ALC	35 Caucasian cirrhotic patients compared with European Caucasian populations. The results show lower $\epsilon 4$ and $\epsilon 2$ allele frequencies and higher $\epsilon 3$ allele frequency in Caucasian alcoholic cirrhotics	Risk of alcoholic cirrhosis.	↑ $\epsilon 3$ ↓ $\epsilon 4 = \epsilon 2$
Hernández-Nazara et al. 2008 [55]	ALC	Case-control study: 86 patients with alcoholic cirrhosis were subdivided in hyperlipidemic and non-hyperlipidemic, and 133 healthy individuals.	Risk of liver cirrhosis: Hyperlipidemic. Non-hyperlipidemic.	↑ $\epsilon 2$ ↓ $\epsilon 4$
Frenzer et al. 2002 [57]	ALC	Case-control study: Caucasian adults (57 with alcoholic cirrhosis, 71 with alcoholic chronic pancreatitis, 57 with alcoholics without apparent organ damage and 200 healthy blood donors).	Risk of alcoholic cirrhosis.	↑ $\epsilon 3/\epsilon 3$ ↓ $\epsilon 4/\epsilon 4$
De Feo et al. 2012 [63]	NAFLD	310 cases and 422 controls were genotyped for apoE.	NAFLD risk.	↑ $\epsilon 3$ ↓ $\epsilon 4$
Sazci et al. 2008 [64]	NASH	57 cases and 245 controls were genotyped for apoE.	NASH risk.	↑ $\epsilon 3$ , $\epsilon 3/\epsilon 3$ ↓ $\epsilon 4$ , $\epsilon 2/\epsilon 3$ and $\epsilon 3/\epsilon 4$
Demirag et al. 2007 [65]	NAFLD	237 NAFLD patients and 201 controls in Turkish population.	Protective against NAFLD.	↑ $\epsilon 2$ , $\epsilon 2/\epsilon 3$
Yang et al. 2005 [66]	NAFLD	Cross-sectional study: 711 patients with NAFLD and 711 control subjects.	NAFLD risk.	↑ $\epsilon 4$
Stachowska et al. 2013 [67]	NAFLD	Prospective study: 23 patients with NAFLD, 11 patients in the apoE4 group were compared with 12 in the apoE3 group.	Risk of advanced fibrosis.	↑ $\epsilon 4$ ↓ $\epsilon 3$

Hepatitis C virus (HCV); Normal alanine aminotransferase levels (PNALT); Hepatocellular carcinoma (HCC); Hepatitis B virus (HBV); Primary biliary cirrhosis (PBC); Alcoholic liver cirrhosis (ALC); Non-alcoholic fatty liver disease (NAFLD); Increased allele frequency (↑); Decrease allele frequency (↓); Apolipoprotein (apoE).

a reduction in the frequency of lipid-carrying proteins, resulting in a decrease in the frequency of apolipoproteins in plasma [56].

In the analysis of patients with hyperlipidaemic or non-hyperlipidaemic alcoholic liver cirrhosis, Hernández-Nazara et al. [55] observed that the  $\epsilon 2$  allele was closely associated with the hyperlipidaemic group and the early onset of cirrhosis, with alcohol intake <20 years, and that the allele  $\epsilon 4$  was more frequent in the non-hyperlipidaemic group and apoE  $\epsilon 4$  carriers were more

resistant to cirrhosis with alcohol consumption >20 years. Iron et al. [54] studied alcoholic cirrhotic patients and observed that there was a higher frequency of  $\epsilon 4$  and  $\epsilon 2$  alleles. Frenzer et al. [57] did not identify a significant difference in apoE genotype frequencies when compared to alcoholic cirrhosis, pancreatitis, controls and blood donor groups. Furthermore, in the cirrhotic group, the  $\epsilon 3/\epsilon 3$  genotype had a higher frequency, whereas  $\epsilon 4/\epsilon 4$  was the least frequent.

## 7. Non-alcoholic fatty liver disease or non-alcoholic steatohepatitis

Non-alcoholic fatty liver disease (NAFLD) is characterised as the accumulation of excess liver fat and increased endogenous lipogenesis substances unrelated to chronic alcoholism [58,59]. Most of these patients may present with metabolic alterations such as dyslipidaemia, diabetes mellitus and obesity. In addition, they may develop non-alcoholic steatohepatitis (NASH), which may progress to fibrosis and cirrhosis [58,60]. ApoE is associated with different pathologies as well as with altered lipid profiles [61]. Previous studies with apoE deficient mice fed a cholesterol-rich diet have shown that this method may be a valuable alternative in NASH research [62]. Interestingly,  $\epsilon 4$  allele patients had a significantly lower risk of NAFLD and a significant reduction in HDL cholesterol [63].

In another study, the  $\epsilon 3/\epsilon 3$  genotype and  $\epsilon 3$  allele were more prevalent in patients with NAFLD and also associated with increased risk of NASH, whereas the  $\epsilon 2/\epsilon 3$  and  $\epsilon 3/\epsilon 4$  genotypes and allele  $\epsilon 4$  were associated with protection against NASH [64]. In addition, the  $\epsilon 2$  allele and  $\epsilon 2/\epsilon 3$  genotype have been shown protective against the development of NAFLD [65]. Conversely, other studies suggest that the apoE  $\epsilon 4$  allele is a risk factor for NAFLD pathogenesis [66].

## 8. Role of apolipoprotein E in liver fibrosis

A protective effect of apoE  $\epsilon 4$  against severe hepatic fibrosis has been supported by previous findings of lower  $\epsilon 4$  allele frequency among patients with HCV-related severe hepatic fibrosis compared to those with mild liver disease [24]. Fabris et al. have reported a benefit of the apoE  $\epsilon 4$  allele on the progression of fibrosis in liver transplant patients with recurrent hepatitis C [19]. However, in another study by Stachowska et al. [67] evaluating NAFLD, the  $\epsilon 4$  allele was significantly associated with the development of advanced fibrosis due to disrupted hepatic fatty acid metabolism and increased 5-oxo-6,8,11,14-eicosatetraenoic acid production [66]. In addition, a study by Mueller et al. could not find an  $\epsilon 4$  allele-protective effect against liver fibrosis in patients diagnosed with chronic HCV infection [29]. Nonetheless, the  $\epsilon 3/\epsilon 3$  occurrence has been correlated with the rapid progression of fibrosis [27].

ApoE can facilitate cholesterol efflux from peripheral macrophages and other cells to form nascent discoidal high-density lipoprotein (HDL) after interaction with ABC transporters. The HDL particles deliver the cholesterol to hepatocytes via interaction with scavenger receptor B1 (SR-B1) or low-density lipoprotein (LDL) receptors. There is later conversion to cholesterol-bearing bile acids, therefore contributing to the reverse cholesterol transport from periphery to hepatocytes and to faecal excretion [68].

Hepatocytes can become steatotic by prolonged high-fat diets, indicating that hepatocytes may be overloaded by excess cholesterol delivery. Increased lipolysis (decreased  $\beta$ -oxidation of triglycerides) or increased de novo lipogenesis and reduced VLDL secretion from hepatocytes are also involved in this process [69]. ApoE has been recognised as an important contributing factor to improving liver steatosis. It has been shown that apoE-deficient mice chronically fed with western diets develop non-alcoholic steatohepatitis and liver fibrosis [62].

Chronic liver inflammation and liver steatosis may increase the rates of hepatocyte apoptosis and significantly increase the expression of platelet-derived growth factor BB (PDGF-BB) and transforming growth factor  $\beta$  (TGF $\beta$ ) that lead to activation of Ito cells (hepatic stellate cells) and pro-fibrogenesis-activated genes [70]. Ito cells, which represent quiescent liver vitamin A-storing

cells in the physiological state, are activated during chronic liver inflammation. These cells are transdifferentiated to myofibroblasts that produce extracellular matrix collagens and a myriad of inflammatory signals that lead to liver fibrosis and loss of vitamin A storage [71]. Interestingly, cultured Ito cells are able to synthesise and release apoE peptides [72].

It remains elusive whether apoE  $\epsilon 4$  could be released during fibrosis and whether this peptide could improve this process.

Table 1 summarises studies addressing apoE polymorphisms and liver diseases in clinical studies to date.

## 9. Conclusion

Overall, this concise review has summarised accumulating evidence that apoE  $\epsilon 4$  carriers are protected against chronic HCV infection, have slow progression of liver fibrosis, and are less likely to have alcoholic cirrhosis, NASH, HCC or HBV. On the other hand, apoE  $\epsilon 3$  carriers are at higher risk for developing liver cirrhosis caused by NASH, NAFLD, HCC or HBV. We still can find contradictory results regarding the protective role of apoE  $\epsilon 4$  on liver fibrosis and NAFLD. In addition, a gap of knowledge still exists regarding the apoE genotypes' role on the mechanisms of liver injury following viral hepatitis. Interactional mechanisms of apoE with hepatitis viruses may be a potential target for gene-related therapies in the future.

Although further clinical studies are highly needed, in this essential review, we highlight that carriage of the apoE  $\epsilon 4$  allele may exert a protective effect in reducing the progression of most liver diseases from different aetiologies.

### Abbreviations

HCV	hepatitis C virus
ApoE	apolipoprotein E
NASH	non-alcoholic steatohepatitis
PBC	primary biliary cirrhosis
VLDL	very low density lipoprotein
LDL-R	low density lipoprotein receptor
IL-2	interleukin-2
HIV	human immunodeficiency virus
HDL	high density lipoprotein
LDL	low density lipoprotein
CSF	cerebrospinal fluid
HBV	hepatitis B virus
HCC	hepatocellular carcinoma
HSV	herpes simplex virus
IL-6	interleukin-6
HSPGs	heparan sulfate proteoglycans
LVP	lipoviral particle
PBC	primary biliary cirrhosis
NAFLD	non-alcoholic fatty liver disease
SR-B1	scavenger receptor-B1
PDGF-BB	platelet-derived growth factor-BB
TGF $\beta$	transforming growth factor $\beta$

### Contribution of authors

J.C.R.N., G.A.M., A.E.C.C.B.M, L.C.P., A.M.S., R.B.O., P.T. contributed to the study design and writing of the manuscript; J.C.R.N., R.B.O., P.T. critically revised the manuscript for important intellectual content and supervised the study.

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### Conflicts of interest

There are no conflicts of interest.

### References

- [1] Iredale JP. Cirrhosis: new research provides a basis for rational and targeted treatments. *BMJ* 2003;327:143–7.
- [2] Martínez-Esparza M, Tristan-Manzano M, Ruiz-Alcaraz AJ, García-Penarrubia P. Inflammatory status in human hepatic cirrhosis. *World J Gastroenterol* 2015;21:11522–41.
- [3] Mas VR, Fassnacht R, Archer KJ, Maluf D. Molecular mechanisms involved in the interaction effects of alcohol and hepatitis C virus in liver cirrhosis. *Mol Med* 2010;16:287–97.
- [4] Mueller S, Millonig G, Seitz HK. Alcoholic liver disease and hepatitis C: a frequently underestimated combination. *World J Gastroenterol* 2009;15:3462–71.
- [5] Zannis VI, Breslow JL, Utermann G, Mahley RW, Weisgraber KH, Havel RJ, et al. Proposed nomenclature of apoE isoproteins, apoE genotypes, and phenotypes. *J Lipid Res* 1982;23:911–4.
- [6] Mahley RW, Huang Y, Rall Jr SC. Pathogenesis of type III hyperlipoproteinemia (dysbetalipoproteinemia), questions, quandaries, and paradoxes. *J Lipid Res* 1999;40:1933–49.
- [7] Mahley RW, Rall Jr SC. Apolipoprotein E: far more than a lipid transport protein. *Annu Rev Genomics Hum Genet* 2000;1:507–37.
- [8] Driscoll DM, Getz GS. Extrahepatic synthesis of apolipoprotein E. *J Lipid Res* 1984;25:1368–79.
- [9] Kraft HG, Menzel HJ, Hoppichler F, Vogel W, Utermann G. Changes of genetic apolipoprotein phenotypes caused by liver transplantation. Implications for apolipoprotein synthesis. *J Clin Invest* 1989;83:137–42.
- [10] Tudorache IF, Trusca VG, Gafencu AV. Apolipoprotein E – a multifunctional protein with implications in various pathologies as a result of its structural features. *Comput Struct Biotechnol J* 2017;15:359–65.
- [11] Schmitz F, Mevissen V, Krantz C, Kimmel M, Erdmann J, Hoffmann R, et al. Robust association of the APOE epsilon4 allele with premature myocardial infarction especially in patients without hypercholesterolemia: the Aachen study. *Eur J Clin Invest* 2007;37:106–8.
- [12] Oriá RB, Patrick PD, Blackman JA, Lima AA, Guerrant RL. Role of apolipoprotein E4 in protecting children against early childhood diarrhea outcomes and implications for later development. *Med Hypotheses* 2007;68:1099–107.
- [13] Jorns C, Takahashi T, Callaghan E, Zemack H, Larsson L, Nowak G, et al. Serum apolipoprotein E as a marker to monitor graft function after hepatocyte transplantation in a clinically relevant mouse model. *Transplant Proc* 2013;45:1780–6.
- [14] Utermann G, Langenbeck U, Beisiegel U, Weber W. Genetics of the apolipoprotein E system in man. *Am J Hum Genet* 1980;32:339–47.
- [15] Linton MF, Gish R, Hubl ST, Büttler E, Esquivel C, Bry WI, et al. Phenotypes of apolipoprotein B and apolipoprotein E after liver transplantation. *J Clin Invest* 1991;88:270–81.
- [16] Eichner JE, Dunn ST, Perveen G, Thompson DM, Stewart KE, Stroehla BC. Apolipoprotein E polymorphism and cardiovascular disease: a HuGE review. *Am J Epidemiol* 2002;155:487–95.
- [17] Koopal C, Marais AD, Westerink J, Visseren FL. Autosomal dominant familial dysbetalipoproteinemia: a pathophysiological framework and practical approach to diagnosis and therapy. *J Clin Lipidol* 2017;11:12–23.
- [18] Schneider WJ, Kovanen PT, Brown MS, Goldstein JL, Utermann G, Weber W, et al. Familial dysbetalipoproteinemia. Abnormal binding of mutant apoprotein E to low density lipoprotein receptors of human fibroblasts and membranes from liver and adrenal of rats, rabbits, and cows. *J Clin Invest* 1981;68:1075–85.
- [19] Fabris C, Toniutto P, Bitetto D, Minisini R, Smirne C, Caldato M, et al. Low fibrosis progression of recurrent hepatitis C in apolipoprotein E epsilon4 carriers: relationship with the blood lipid profile. *Liver Int* 2005;25:1128–35.
- [20] Yin Z, Xiong C, Wang Y, Zhou X, Yan SK. Investigation of the relationship between apolipoprotein E gene polymorphisms and hepatitis B virus infection in northern China. *Clin Chem Lab Med* 2010;48:1803–7.
- [21] Shen Y, Li M, Ye X, Bi Q. Association of apolipoprotein E with the progression of hepatitis B virus-related liver disease. *Int J Clin Exp Pathol* 2015;8:14749–56.
- [22] Franco E, Bagnato B, Marino MG, Meleleo C, Serino L, Zaratti L, et al. Epidemiology and prevention in developing countries. *World J Hepatol* 2012;4:74–80.
- [23] Kuhlmann I, Minihane AM, Huebbe P, Nebel A, Rimbach G. Apolipoprotein E genotype and hepatitis C, HIV and herpes simplex disease risk: a literature review. *Lipids Health Dis* 2010;9:8.
- [24] Wozniak MA, Itzhaki RF, Faragher EB, James MW, Ryder SD, Irving WL. Apolipoprotein E-epsilon4 protects against severe liver disease caused by hepatitis C virus. *Hepatology* 2002;36:456–63.
- [25] Ahn SJ, Kim DK, Kim SS, Bae CB, Cho HJ, Kim HG, et al. Association between apolipoprotein E genotype, chronic liver disease, and hepatitis B virus. *Clin Mol Hepatol* 2012;18:295–301.
- [26] Choo QL, Kuo G, Weiner AJ, Overby LR, Bradley DW, Houghton M. Isolation of a cDNA clone derived from a blood-borne non-A, non-B viral hepatitis genome. *Science* 1989;244:359–62.
- [27] Fabris C, Vandelli C, Toniutto P, Minisini R, Colletta C, Falleti E, et al. Apolipoprotein E genotypes modulate fibrosis progression in patients with chronic hepatitis C and persistently normal transaminases. *J Gastroenterol Hepatol* 2011;26:328–33.
- [28] Hishiki T, Shimizu Y, Tobita R, Sugiyama K, Ogawa K, Funami K, et al. Infectivity of hepatitis C virus is influenced by association with apolipoprotein E isoforms. *J Virol* 2010;84:12048–57.
- [29] Mueller T, Gessner R, Sarrazin C, Graf C, Halangj J, Witt H, et al. Apolipoprotein E4 allele is associated with poor treatment response in hepatitis C virus (HCV) genotype 1. *Hepatology* 2003;38:1592–3.
- [30] Price DA, Bassendine MF, Norris SM, Golding C, Toms GL, Schmid ML, et al. Apolipoprotein epsilon3 allele is associated with persistent hepatitis C virus infection. *Gut* 2006;55:715–8.
- [31] Mueller T, Fischer J, Gessner R, Rosendahl J, Böhm S, van Bömmel F, et al. Apolipoprotein E allele frequencies in chronic and self-limited hepatitis C suggest a protective effect of APOE4 in the course of hepatitis C virus infection. *Liver Int* 2016;36:1267–74.
- [32] Teama SHH, Agwa S, Makhlof M, Nashaat E, Sayed M, Yousry W, et al. Apolipoprotein-E gene polymorphism and possible role of ApoE e4 allele with a lower probability of progression to HCV-related liver cirrhosis in Egyptian patients. *Merit Res J Med Sci* 2016;4:440–7.
- [33] Wozniak MA, Lugo Iparraquirre LM, Dirks M, Deb-Chatterji M, Pflugrad H, Goldbecker A, et al. Apolipoprotein E-epsilon4 deficiency and cognitive function in hepatitis C virus-infected patients. *J Viral Hepat* 2016;23:39–46.
- [34] Weller R, Hueging K, Brown RJP, Todt D, Joecks S, Vondran FWR, et al. Hepatitis C virus strain-dependent usage of apolipoprotein E modulates assembly efficiency and specific infectivity of secreted virions. *J Virol* 2017;91:e00422–e517.
- [35] Popescu CI, Dubuisson J. Role of lipid metabolism in hepatitis C virus assembly and entry. *Biol Cell* 2009;102:63–74.
- [36] André P, Komurian-Pradel F, Deforges S, Perret M, Berland JL, Sodoier M, et al. Characterization of low- and very-low-density hepatitis C virus RNA-containing particles. *J Virol* 2002;76:6919–28.
- [37] Felmlée DJ, Hafirassou ML, Lefevre M, Baumert TF, Schuster C. Hepatitis C virus, cholesterol and lipoproteins – impact for the viral life cycle and pathogenesis of liver disease. *Viruses* 2013;5:1292–324.
- [38] Hueging K, Doeppke M, Vieyres G, Bankwitz D, Frentzen A, Doerrbecker J, et al. Apolipoprotein E codetermines tissue tropism of hepatitis C virus and is crucial for viral cell-to-cell transmission by contributing to a postenvelopment step of assembly. *J Virol* 2014;88:1433–46.
- [39] Yang Z, Wang X, Chi X, Zhao F, Guo J, Ma P, et al. Neglected but important role of apolipoprotein E exchange in hepatitis C virus infection. *J Virol* 2016;90:9632–43.
- [40] Monazahian M, Böhme I, Bonk S, Koch A, Scholz C, Grethe S, et al. Low density lipoprotein receptor as a candidate receptor for hepatitis C virus. *J Med Virol* 1999;57:223–9.
- [41] Tinkle CL, Haas-Kogan D. Hepatocellular carcinoma: natural history, current management, and emerging tools. *Biologics* 2012;6:207–19.
- [42] Bruix J, Sherman M. Management of hepatocellular carcinoma: an update. *Hepatology* 2011;53:1020–2.
- [43] Mazzaferro V, Regalia E, Doci R, Andreola S, Pulvirenti A, Bozzetti F, et al. Liver transplantation for the treatment of small hepatocellular carcinomas in patients with cirrhosis. *N Engl J Med* 1996;334:693–9.
- [44] Jensen PR, Serra SC, Miragoli L, Karlsson M, Cabella C, Poggi L, et al. Hyperpolarized [1,3-<sup>13</sup>C]ethyl acetoacetate is a novel diagnostic metabolic marker of liver cancer. *Int J Cancer* 2015;136:E117–26.
- [45] Oue N, Hamai Y, Mitani Y, Matsumura S, Oshimo Y, Aung PP, et al. Gene expression profile of gastric carcinoma: identification of genes and tags potentially involved in invasion, metastasis, and carcinogenesis by serial analysis of gene expression. *Cancer Res* 2004;64:2397–405.
- [46] Chen YC, Pohl G, Wang TL, Morin PJ, Risberg B, Kristensen GB, et al. Apolipoprotein E is required for cell proliferation and survival in ovarian cancer. *Cancer Res* 2005;65:331–7.
- [47] Liu DY, Peng ZH, Qiu GQ, Zhou CZ. Expression of telomerase activity and oxidative stress in human hepatocellular carcinoma with cirrhosis. *World J Gastroenterol* 2003;9:1859–62.
- [48] Yokoyama Y, Kuramitsu Y, Takashima M, Iizuka N, Terai S, Oka M, et al. Protein level of apolipoprotein E increased in human hepatocellular carcinoma. *Int J Oncol* 2006;28:625–31.
- [49] Hirschfield GM, Gershwin ME. The immunobiology and pathophysiology of primary biliary cirrhosis. *Annu Rev Pathol* 2013;8:303–30.
- [50] Corpechot C, Benlian P, Barbu V, Chazouilleres O, Pouyon RE, Pouyon R. Apolipoprotein E polymorphism, a marker of disease severity in primary biliary cirrhosis? *J Hepatol* 2001;35:324–8.
- [51] Vuoristo M, Färkkilä M, Gylling H, Karvonen AL, Leino R, Lehtola J, et al. Expression and therapeutic response related to apolipoprotein E polymorphism in primary biliary cirrhosis. *J Hepatol* 1997;27:136–42.
- [52] Kerr WC, Fillmore KM, Marvy P. Beverage-specific alcohol consumption and cirrhosis mortality in a group of English-speaking beer-drinking countries. *Addiction* 2000;95:339–46.
- [53] Rehm J, Taylor B, Mohapatra S, Irving H, Baliunas D, Patra J, et al. Alcohol as a risk factor for liver cirrhosis: a systematic review and meta-analysis. *Drug Alcohol Rev* 2010;29:437–45.

- [54] Iron A, Richard P, Pascual De ZM, Dumas F, Cassaigne A, Couzigou P. Genetic polymorphism of apolipoprotein E in Caucasian alcoholic cirrhotics. *Alcohol Alcohol* 1994;29:715–8.
- [55] Hernandez-Nazara ZH, Ruiz-Madrigal B, Martinez-Lopez E, Roman S, Panduro A. Association of the epsilon 2 allele of APOE gene to hypertriglyceridemia and to early-onset alcoholic cirrhosis. *Alcohol Clin Exp Res* 2008;32:559–66.
- [56] Bhopale KK, Amer SM, Kaphalia L, Soman KV, Wiktorowicz JE, Shakeel Ansari GA, et al. Proteomic profiling of liver and plasma in chronic ethanol feeding model of hepatic alcohol dehydrogenase-deficient deer mice alcohol. *Clin Exp Res* 2017;41:1675–85.
- [57] Frenzer A, Butler WJ, Norton ID, Wilson JS, Apte MV, Pirola RC, et al. Polymorphism in alcohol-metabolizing enzymes, glutathione S-transferases and apolipoprotein E and susceptibility to alcohol-induced cirrhosis and chronic pancreatitis. *J Gastroenterol Hepatol* 2002;17:177–82.
- [58] Chalasani N, Younossi Z, Lavine JE, Charlton M, Cusi K, Rinella M, et al. The diagnosis and management of nonalcoholic fatty liver disease: practice guidance from the American Association for the Study of Liver Diseases. *Hepatology* 2018;67:328–57.
- [59] Gentile CL, Pagliassotti MJ. The role of fatty acids in the development and progression of nonalcoholic fatty liver disease. *J Nutr Biochem* 2008;19:567–76.
- [60] Puri P, Baillie RA, Wiest MM, Mirshahi F, Choudhury J, Cheung O, et al. A lipidomic analysis of nonalcoholic fatty liver disease. *Hepatology* 2007;46:1081–90.
- [61] Papaioannou I, Simons JP, Owen JS. Targeted in situ gene correction of dysfunctional APOE alleles to produce atheroprotective plasma ApoE3 protein. *Cardiol Res Pract* 2012;2012:148796.
- [62] Schierwagen R, Maybüchen L, Zimmer S, Hittatiya K, Bäck C, Klein S, et al. Seven weeks of Western diet in apolipoprotein-E-deficient mice induce metabolic syndrome and non-alcoholic steatohepatitis with liver fibrosis. *Clin Rep* 2015;5:12931.
- [63] De Feo E, Cefalo C, Arzani D, Amore R, Landolfi R, Grieco A, et al. A case-control study on the effects of the apolipoprotein E genotypes in nonalcoholic fatty liver disease. *Mol Biol Rep* 2012;39:7381–8.
- [64] Sazci A, Akpinar G, Aygun C, Ergul E, Senturk O, Hulagu S. Association of apolipoprotein E polymorphisms in patients with non-alcoholic steatohepatitis. *Dig Dis Sci* 2008;53:3218–24.
- [65] Demirag MD, Onen HI, Karaoguz MY, Dogan I, Karakan T, Ekmekci A, et al. Apolipoprotein E gene polymorphism in nonalcoholic fatty liver disease. *Dig Dis Sci* 2007;52:3399–403.
- [66] Yang MH, Son HJ, Sung JD, Choi YH, Koh KC, Yoo BC, et al. The relationship between apolipoprotein E polymorphism, lipoprotein (a) and fatty liver disease. *Hepatogastroenterology* 2005;52:1832–5.
- [67] Stachowska E, Maciejewska D, Ossowski P, Drozd A, Ryterska K, Banaszczak M, et al. Apolipoprotein E4 allele is associated with substantial changes in the plasma lipids and hyaluronic acid content in patients with nonalcoholic fatty liver disease. *J Physiol Pharmacol* 2013;64:711–7.
- [68] Getz GS, Reardon CA. Apoprotein E and reverse cholesterol transport. *Int J Mol Sci* 2018;19, pii:E3479.
- [69] Koyama Y, Brenner DA. Liver inflammation and fibrosis. *J Clin Invest* 2017;127:55–64.
- [70] Mekala S, Tulimilli SV, Geesala R, Manupati K, Dhoke NR, Das A. Cellular crosstalk mediated by platelet-derived growth factor BB and transforming growth factor  $\beta$  during hepatic injury activates hepatic stellate cells. *Can J Physiol Pharmacol* 2018;96:728–41.
- [71] Tsuchida T, Friedman SL. Mechanisms of hepatic stellate cell activation. *Ann Gastroenterol Surg* 2017;1:52–9.
- [72] Ramadori G, Rieder H, Theiss F, Meyer zum Büschenfelde KH. Fat-storing (Ito) cells of rat liver synthesize and secrete apolipoproteins: comparison with hepatocytes. *Gastroenterology* 1989;97:163–72.